CMV/EBV/HHV6 Quant Real-TM

Handbook

Multiplex Real Time PCR Kit for quantitative detection and differentiation of Cytomegalovirus (CMV), Epstein Barr Virus (EBV) and Human Herpes 6 Virus (HHV6)

REF TV48-100FRT

100

NAME

CMV/EBV/HHV6 Quant Real - TM

INTENDED USE

The **CMV/EBV/HHV6 Quant Real-TM** is a “Real-Time Amplification” test for the quantitative detection and differentiation of Cytomegalovirus (CMV), Epstein Barr Virus (EBV) and Human Herpes 6 Virus (HHV6) in the biological materials. DNA is extracted from samples, amplified using real time amplification with fluorescent reporter dye probes specific for CMV/EBV/HHV6 and Internal Control (IC). Test contains an IC (β-globine gene) which allows controlling both PCR-analysis stages (DNA extraction and PCR amplification), material sampling, and storage conditions.

PRINCIPLE OF PCR DETECTION

CMV, EBV and HHV6 detection by polymerase chain reaction (PCR) with hybridization-fluorescence detection includes DNA extraction from clinical samples and PCR amplification of pathogen genome specific region with real-time hybridization-fluorescence detection. During DNA extraction from clinical material, human genomic DNA (endogenous internal control) is amplified. Endogenous internal control (IC Glob) allows controlling both PCR-analysis stages (DNA extraction and PCR amplification), material sampling, and storage adequacy. Then, the obtained samples are amplified using specific primers and polymerase (TaqF). In real-time PCR, the amplified product is detected using fluorescent dyes. These dyes are linked to oligonucleotide probes which bind specifically to the amplified product during thermocycling. The real-time monitoring of the fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

MATERIALS PROVIDED

Part N° 1 – **"DNA-Sorb-B":** isolation of DNA from clinical specimens

- **Lysis Solution**, 2 x 15 ml;
- **Washing Solution 1**, 2 x 15 ml;
- **Washing Solution 2**, 2 x 50 ml;
- **Sorbent**, 2 x 1,25 ml;
- **DNA-eluent**, 2 x 5,0 ml.

Contains reagents for 100 extractions

Part N° 2– **"CMV/EBV/HHV6 Quant Real-TM":** Real Time Amplification;

- **PCR-mix-1-FRT *EBV / CMV / HHV-6* / Glob,** 2 x 0,6 ml;
- **PCR-mix-2-FRT**, 2 x 0,3 ml;
- **Polymerase (TaqF),** 2 x 0,03 ml;
- **RNA-buffer**, 0,6 ml
- **Negative Control C-***, 2 x 1,2 ml;
- **Positive Control DNA *EBV / CMV / HHV-6* and human DNA**,** 2 x 0,1 ml;
- **Standard CMV/EBV/HHV6//IC glob**
    - **DNA calibrator ĸSG1,** 0,2 ml;
    - **DNA calibrator ĸSG2**, 0,2 ml;

Contains reagents for 120 tests.

* *must be used in the extraction procedure as Negative Control of Extraction.*
** *must be used in the extraction procedure as Positive Control of Extraction (PCE).*

MATERIALS REQUIRED BUT NOT PROVIDED

Zone 1: sample preparation:

- Biological cabinet
- Desktop microcentrifuge for “eppendorf” type tubes
- 60°C ± 5°C dry heat block
- Vortex mixer
- Pipettes (adjustable)
- 1,5 ml polypropylene sterile tubes
- Disposable gloves, powderless
- Biohazard waste container
- Refrigerator, Freezer

Zone 2: RT and amplification:

- Desktop microcentrifuge for “eppendorf” type tubes
- Vortex mixer
- Disposable gloves, powderless
- Biohazard waste container
- Refrigerator, Freezer
- Real Time Thermal cycler
- Workstation
- Pipettes (adjustable)
- Sterile pipette tips with filters

STORAGE INSTRUCTIONS

CMV/EBV/HHV6 Quant Real-TM must be stored at - 20°C. The kit can be shipped at 2-8°C but should be stored at -20°C immediately on receipt. **DNA-Sorb-B** kit can be stored at room temperature.

STABILITY

CMV/EBV/HHV6 Quant Real-TM test is stable up to the expiration date indicated on the kit label. The product will maintain performance through the control date printed on the label. Exposure to light, heat or humidity may affect the shelf life of some of the kit components and should be avoided. Repeated thawing and freezing of these reagents should be avoided, as this may reduce the sensitivity. Components stored under conditions other than those stated on the labels may not perform properly and may adversely affect the assay results.

WARNINGS AND PRECAUTIONS

The user should always pay attention to the following:

- ⚠ Lysis Solution contains guanidine thiocyanate*. Guanidine thiocyanate is harmful if inhaled, or comes into contact with skin or if swallowed. Contact with acid releases toxic gas. (Xn; R: 20/21/22-36/37/38; S: 36/37/39).
- Use sterile pipette tips with aerosol barriers and use new tip for every procedure.
- Store extracted positive material (samples, controls and amplicons) away from all other reagents and add it to the reaction mix in a separate area.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable gloves, laboratory coats and eye protection when handling specimens and reagents. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local authorities' regulations.
- Specimens should be considered potentially infectious and handled in a biological cabinet in accordance with appropriate biosafety practices.
- Clean and disinfect all sample or reagent spills using a disinfectant such as 0.5% sodium hypochlorite, or other suitable disinfectant.
- Avoid sample or reagent contact with the skin, eyes, and mucous membranes. If skin, eyes, or mucous membranes come into contact, rinse immediately with water and seek medical advice immediately.
- Material Safety Data Sheets (MSDS) are available on request.
- Use of this product should be limited to personnel trained in the techniques of DNA amplification.
- The laboratory process must be one-directional, it should begin in the Extraction Area and then move to the Amplification and Detection Areas. Do not return samples, equipment and reagents to the area in which the previous step was performed.

Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

PRODUCT USE LIMITATIONS

Use of this product should be limited to personnel trained in the techniques of DNA amplification (UNI EN ISO 18113-2:2012). Strict compliance with the user manual is required for optimal PCR results. Attention should be paid to expiration dates printed on the box and labels of all components. Do not use a kit after its expiration date.

SAMPLE COLLECTION, STORAGE AND TRANSPORT

CMV/EBV/HHV6 Quant Real-TM can analyze DNA extracted from:

- *whole peripheral and umbilical cord blood* collected in either ACD or EDTA tubes;
- *buffy coat*;
- *plasma*;
- *tissue* homogenized with mechanical homogenizer and dissolved in PBS sterile;
- *urine (sediment)*;
- *swabs:* insert the swab into the nuclease-free 1,5 ml tube and add 0,2 mL of Transport medium. Vigorously agitate swabs in medium for 15-20 sec.
- *CSF (Liquor);*

It is recommended to process samples immediately after collection. Store samples at 2–8 °C for no longer than 24 hours, or freeze at –20/80°C. Transportation of clinical specimens must comply with country, federal, state and local regulations for the transport of etiologic agents.

DNA ISOLATION

The following isolation kit is recommended:

⇒ **DNA-Sorb-B** (Sacace, REF K-1-1/B)

⇒ **DNA/RNA-Prep** (Sacace, REF K-2-9)

⇒ **SaMag Viral Nucleic Acids Extraction Kit** (Sacace, REF SM003, for plasma)

⇒ **SaMag STD DNA Extraction Kit** (Sacace, REF SM007, for urine sediment)

Extract DNA according to the manufacturer's instructions.

Transfer **100µl** of **Negative Control** to the tube labeled C–. Transfer **90µl** of **Negative Control** and**10µl** of **Positive Control DNA *EBV / CMV / HHV-6* and human DNA** to the tube labeled PCE.

SPECIMEN AND REAGENT PREPARATION

1. **Lysis Solution** and **Washing Solution** (in case of their storage at +2-8°C) should be warmed up to 60°C until disappearance of ice crystals.
2. Prepare required quantity of 1.5 ml polypropylene tubes.
3. Add to each tube **300 µl** of **Lysis Solution**.
4. Add **100 µl** of **Samples** to the appropriate tube.
5. Prepare Controls as follows:
   - add **100 µl** of **C–** (**Negative Control**) to the tube labeled *Cneg extraction*.
   - add **10 µl** of **Positive Control DNA *EBV / CMV / HHV-6* and human DNA** and **90 µl** of **C–** (**Negative Control**) to the tube labeled PCE
6. Vortex the tubes, incubate 5 min at 65°C and centrifuge for 5 sec.
7. Vortex vigorously **Sorbent** and add **25 µl** to each tube.
8. Vortex for 5-7 sec and incubate all tubes for 10 min at room temperature. Vortex periodically
9. Centrifuge all tubes for 1 min at 5000g and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between tubes.
10. Add **300 µl** of **Washing Solution 1** to each tube. Vortex vigorously and centrifuge for 1 min at 5000g and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between tubes.
11. Add **500 µl** of **Washing Solution 2** to each tube. Vortex vigorously and centrifuge for 1 min at 10000g and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between tubes.
12. Repeat step 11.
13. Incubate all tubes with open cap for 5 min at 65°C.
16. Resuspend the pellet in **50 µl** of **DNA-eluent.** Incubate for 5 min at 65°C and vortex periodically.
17. Centrifuge the tubes for 2 min at maximum speed (12000-16000 g). The supernatant contains DNA ready for amplification. The amplification can be performed on the same day of extraction.

PROTOCOL (Reaction volume 25 µl):

1. Prepare in the new sterile tube for each sample **10*N µl** of **PCR-mix-1-FRT *EBV/CMV/ HHV-6/*Glob**, **5,0*N** of **PCR-mix-2-FRT** and **0,5*N** of **Polymerase (TaqF).** Vortex and centrifuge for 2-3 sec.
2. Prepare required quantity of reaction tubes for extracted samples (including negative and positive extraction control) and amplification controls and add **15 µl** of **Reaction Mix** and **10 µl** of **extracted DNA** sample to appropriate tube. Mix by pipetting.

(*Re-centrifuge all the tubes with extracted DNA for 2 min at maximum speed (12000-16000 g) and take carefully supernatant. N.B. don't disturb the pellet, sorbent inhibit reaction!).*

3. Prepare for qualitative run 1 tube for negative control and 1 tube for positive control:
   - add **10 µl** of **RNA-buffer** to the tube labeled NCA (Negative Control of Amplification).;
   - add **10 µl** of DNA calibrator **KSG2** to the tube labeled C+ (Positive Control of Amplification);
4. For quantitative analysis prepare 1 tube for negative control and 4 tubes for DNA calibrator KSG1 and KSG2 twice:
   - add **10 µl** of **RNA-buffer** to the tube labeled NCA (Negative Control of Amplification);
   - Add **10 µl** of **KSG1** to two tubes and add **10 µl** of **KSG2** to other two tubes.

Close tubes and transfer them into the instrument in this order: samples, negative controls, positive control, Standards.

Amplification program for rotor-type instruments[1]

Step	*Temperature, °C*	*Time*	*Fluorescence detection*	*Cycles*
Hold	95	15 min	–	1
Cycling 1	95	5 s	–	5
	60	20 s	–	
	72	15 s	–	
Cycling 2	95	5 s	–	40
	60	20 s	FAM/Green, JOE/Yellow, ROX/Orange, Cy5/Red	
	72	15 s	–	

[1] For example Rotor-Gene™ 3000/6000 (Corbett Research, Australia)

Amplification program for plate-type and modular type instruments[2]

Step	*Temperature, °C*	*Time*	*Fluorescence detection*	*Cycles*
1	95	15 min	–	1
2	95	5 s	–	5
	60	20 s	–	
	72	15 s	–	
3	95	5 s	–	40
	60	30 s	FAM, JOE/HEX/Cy3, ROX/TexasRed, Cy5	
	72	15 s	–	

[2] For example, SaCycler-96™ (Sacace), iQ5™/iQ iCycler™ (BioRad, USA); Mx3000P/Mx3005P™ (Stratagene, USA), Applied Biosystems® 7500 Real Time PCR (Applera), SmartCycler® (Cepheid)

RESULTS ANALYSIS

β-Globin gene DNA (IC) is detected in the FAM/Green channel, *EBV* DNA is detected in the JOE/HEX/Cy3/Yellow channel, *CMV* DNA is detected in the ROX/TexasRed/Orange channel, and *HHV6* DNA is detected in the Cy5/Red channel.

Interpretation of results

The results are interpreted by the software of the used Instrument by the crossing (or not) of the fluorescence curve with the threshold line.

1. The sample is considered to be **positive** for *EBV* DNA if its Ct value in the results grid on the JOE/HEX/Cy3/Yellow channel is detected and does not exceed the threshold value of positive result.
2. The sample is considered to be **positive** for *CMV* DNA if its Ct value in the results grid on the ROX/Orange/Texas Red channel is defined and does not exceed the threshold value of positive result.
3. The sample is considered to be **positive** for *HHV6* DNA if its Ct value in the results grid on the Cy5/Red channel is defined and does not exceed the threshold value of positive result.
4. For qualitative analysis, the sample is considered to be **negative** if its Ct value in the results grid in the FAM/Green channel does not exceed the Ct value indicated in the **Boundary Ct values** table.
5. For quantitative analysis, the quantity of IC Glob DNA should be greater than 2000 copies per reaction for whole blood, white blood cells, viscera biopsy material or more than 500 copies per reaction for saliva and oropharyngeal swabs.

*For cerebrospinal fluid (liquor), the Ct value can be greater than the Ct value indicated in the **Boundary Ct values** table in the results grid in the FAM/Green channel or the quantity of IC Glob DNA can be less than 500 copies per reaction in case of quantitative analysis because the cerebrospinal fluid samples may contain a very small number of cells.*

6. For **qualitative** analysis, the result of analysis is considered to be **invalid** if the Ct value is not detected in the results grid (the fluorescence curve does not cross the threshold line) or if it is greater than the threshold value in the JOE/HEX/Yellow, ROX/Orange, or Cy5/Red channel and the Ct value in the results grid in the FAM/Green channel exceeds the Ct value indicated in the **Boundary Ct values** table.
7. For **quantitative** analysis, the analysis result is considered to be **invalid** if the Ct value is not detected in the results grid (the fluorescence curve does not cross the threshold line) or if it is greater than the boundary value in the JOE/Yellow/HEX, ROX/Orange, or Cy5/Red channel and the quantity of IC Glob DNA is less than 2000 copies per reaction for whole blood, white blood cells, viscera biopsy material or if it is less than 500 copies per reaction for saliva and oropharyngeal swabs. In such cases, PCR analysis of the sample should be repeated.
8. For qualitative analysis, results of analysis are considered reliable only if the results obtained for both Positive and Negative Controls of amplification as well as for the Negative Control of extraction are correct. For quantitative analysis, results on C+ should fall in range of concentrations indicated in the **Product Data Card.**

Table. 1. Results for controls

Control	Stage for control	Expected result				Interpretation
		FAM/Green	JOE/HEX/ Cy3/Yellow	ROX/Orange/ Texas Red	Cy5/Red	
NCE	DNA extraction, PCR	Neg	Neg	Neg	Neg	OK
NCA	PCR	Neg	Neg	Neg	Neg	OK
C+	DNA extraction, PCR	POS	POS	POS	POS	OK
QS1 QS2	PCR	Pos	Pos	Pos	Pos	OK

Boundary Ct values for RotorGene 6000/Q instruments (Corbett Research, Qiagen)

Sample	FAM/Green	Joe/HEX/Yellow	ROX/Orange	Cy5/Red
NCE	Absent	Absent	Absent	Absent
NCA	Absent	Absent	Absent	Absent
C+	< 24	< 29	< 29	< 28
QS2	< 26	< 26	< 27	< 27
Test samples	< 23	< 30	< 31	< 31

Boundary Ct values for Plate type instruments like SaCycler-96™ (Sacace); iQ5™ (BioRad); Mx3005P™ (Agilent Technologies); ABI® 7500 Real Time PCR (Applied Biosystems); SmartCycler® (Cepheid)

Sample	FAM/Green	Joe/HEX/Yellow	ROX/Orange	Cy5/Red
NCE	Absent	Absent	Absent	Absent
NCA	Absent	Absent	Absent	Absent
C+	< 29	< 34	< 34	< 33
QS2	< 31	< 31	< 32	< 32
Test samples	< 28	< 35	< 36	< 36

Quantitative results

In quantitative analysis, if total DNA is extracted **from human whole blood, white blood cells** and **biopsy material**, the concentration in log of DNA copies per standard cell quantity (10^5) in control and test samples is calculated by the following formula:

For *CMV*:

$$\log\left\{\frac{\textit{CMV}\text{ DNA copies in PCR sample}}{\text{Glob DNA copies in PCR sample}} \times 2*10^5\right\} = \log\{\textit{CMV}\text{ DNA copies}/10^5\text{ of cells}\}.$$

For *EBV*:

$$\log\left\{\frac{\textit{EBV}\text{ DNA copies in PCR sample}}{\text{Glob DNA copies in PCR sample}} \times 2*10^5\right\} = \log\{\textit{EBV}\text{ DNA copies}/10^5\text{ of cells}\}.$$

For *HHV6*:

$$\log\left\{\frac{\textit{HHV6}\text{ DNA copies in PCR sample}}{\text{Glob DNA copies in PCR sample}} \times 2*10^5\right\} = \log\{\textit{HHV6}\text{ DNA copies}/10^5\text{ of cells}\}.$$

The results can be calculated manually or using Excel tables. To do this copy the names of the samples and insert them in the first column (Column A). Copy the concentrations of EBV DNA from the channel Joe(Yellow)/HEX/Cy3 and paste in the second column of Excel table (Column B). Copy the concentrations of IC Glob from the channel Fam(Green) and paste in the third column of Excel table (Column C). Insert in the column D the formula D=LOG (B/C*200000): log values will appear.

Name	Calc Conc (copies/reaction) Joe(Yellow)/HEX/Cy3	Calc Conc (copies/reaction) Fam(Green)	log EBV/10^5 cells
A	B	C	D
1	8742	125640	4,1
2	253	87787	2,8
3		65765	
4	648	16354	3,9
5		76865	
QS1	9962	9793	
QS1	10011	10143	
QS2	98	103	
QS2	102	97	
Neg PCR			

Use the same procedure for calculation of CMV (ROX/Orange/TexasRed channel) and HHV6 (Cy5/Red channel) log quantity inserting in the column B the relative results.

If total DNA is extracted from **saliva, oropharyngeal swabs** and **cerebrospinal fluid (liquor)**, the concentration of DNA per ml of sample (Conc $_{DNA}$) is calculated by the following formula:

$$\text{Conc}_{DNA} = C_{DNA} \times 100 \text{ (copies/ml)}$$

C_{DNA} is the number of *EBV* DNA copies, or the number of *CMV* DNA copies, or the number of *HHV6* DNA copies in DNA sample.

Table 2. Example of Qualitative Analysis (plate type instrument)

Ct limits (plate type instruments)			
IC	EBV	CMV	HHV6
28	35	36	36

No.	Desription	Fam (IC)	Joe (EBV)	Rox (CMV)	Cy5 (HHV6)	Result	EBV	CMV	HHV6
	Name	Ct	Ct	Ct	Ct				
1	344	27,18			28	*HHV6*	-	-	+
2	445	26,41		34,12	32,1	*CMV, HHV6*	-	+	+
3	451	29,81				*Invalid*	?	?	?
4	456	23,3	28,48		27,7	*EBV, HHV6*	+	-	+
5	461	29,02		35,08		*Invalid-?, (low CMV)*	?	low	?
6	472	24,83	33,28			*EBV*	+	-	-
7	477	17,51	24,06		34,95	*EBV, HHV6*	+	-	+
8	489	21,32	21,85		27,2	*EBV, HHV6*	+	-	+
9	491	23,47	28,15			*EBV*	+	-	-
10	494	29,88				*Invalid*	?	?	?
11	497	16,29	31,06		34,18	*EBV, HHV6*	+	-	+
12	501	18,5		32,64		*CMV*	-	+	-
13	C+	27,23	30,18	28,47	27,25	*OK*			
14	C+	26,06	30,45	27,95	26,58	*OK*			
15	C+	26,37	30,8	28,17	26,73	*OK*			
16	C- (Neg. Control)					*OK*			
17	C- (DNA-buffer)					*OK*			
18	C- (DNA-buffer)					*OK*			

PERFORMANCE CHARACTERISTICS

The kit **CMV/EBV/HHV6 Quant Real-TM** allows to detect *CMV/EBV/HHV6 DNA* in 100% of the tests with a sensitivity of not less than 400 copies/ml.

Specificity of **CMV/EBV/HHV6 Quant Real-TM** PCR kit was confirmed by analysis of reference *CMV* strain AD 169, QCMD panel for *Epstein-Barr virus,* as well as by analysis of clinical material with subsequent confirmation of results by sequencing the amplified fragments. The activity of the PCR kit components with respect to DNA of other viruses (herpes simplex virus types 1 and 2, human herpes virus type 8, Varicella Zoster Virus, Parvovirus B19, and others), bacterial pathogens (*Staphylococcus aureus, Streptococcus pyogenes, Streptococcus agalactiae,* and others) and human DNA was absent. The clinical specificity of **CMV/EBV/HHV6 Quant Real-TM** PCR kit was confirmed in laboratory clinical trials.

Target region: CMV – MIE, EBV – LMP, HHV6 – pol gene

TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

1. The presence of any Ct value on JOE/Yellow/HEX, FAM/Green, ROX/Orange and Cy5/Red channels in the results grid for the Negative Control of Amplification (NCA) and for the Neg. Control of Extraction (C-) indicates contamination of reagents or samples. In this case, PCR analysis should be repeated for all samples in which pathogen DNA was detected starting from the DNA extraction stage.
2. For qualitative analysis, if the Ct value in the results grid for the Positive Control of PCR on the JOE/Yellow/HEX, FAM/Green, ROX/Orange, or Cy5/Red channels is absent, it is necessary to repeat amplification for all samples where pathogen DNA was not detected.
3. If the Ct value for the sample is not detected on JOE/Yellow/HEX/Cy3, ROX/Orange/TexasRed, Cy5/Red channel or it exceeds the boundary Ct value specified in the Data Sheet and the Ct value for the sample is greater than the maximum Ct value for IC in the FAM/Green channel, analysis should be repeated starting from the DNA extraction stage. This error may be caused by incorrect treatment of clinical material, which resulted in the loss of DNA, or by the presence of PCR inhibitors.
4. If the Ct value for the sample is detected in JOE/Yellow/HEX/Cy3, ROX/Orange/TexasRed or Cy5/Red channel and it is greater than the boundary Ct value specified in the Data Sheet**,** the result is considered to be equivocal. It is necessary to repeat analysis of such sample in duplicate. If a reproducible positive Ct value is obtained, the result is considered to be positive; otherwise, the result is considered to be equivocal.

KEY TO SYMBOLS USED

REF	List Number
LOT	Lot Number
	Expiration Date
	Store at
	Manufacturer
	Consult instructions for use

	Caution!
	Contains sufficient for <n> tests
VER	Version
NCA	Negative Control of Amplification
NCE	Negative control of Extraction
C+	Positive Control of Amplification
IC	Internal Control

*iQ5™ is a registered trademark of Bio-Rad Laboratories
* Rotor-Gene™ Technology is a registered trademark of Qiagen
* MX3005P® is a registered trademark of Agilent Technologies
*ABI® is a registered trademark of Applied Biosystems
* SaCycler™ is a registered trademark of Sacace Biotechnologies


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