



For in Vitro Diagnostic Use



# VZV Real-TM

# Handbook

Real Time PCR kit for qualitative detection of Varicella Zoster virus (VZV)

REF V61-50FRT

REF TV61-50FRT

∑∕ 50

#### **NAME**

#### **VZV Real-TM**

#### INTRODUCTION

Varicella Zoster virus (VZV) is known by many names, including: chickenpox virus, varicella virus, zoster virus, and human herpes virus type 3 (HHV-3). Primary VZV infection results in chickenpox (varicella), which may rarely result in complications including encephalitis or pneumonia. Even when clinical symptoms of chickenpox have resolved, VZV remains dormant in the nervous system of the infected person (virus latency), in the trigeminal and dorsal root ganglia. In about 10-20% of cases, VZV reactivates later in life producing a disease known as herpes zoster or shingles. Serious complications of shingles include postherpetic neuralgia, zoster multiplex, myelitis, herpes ophthalmicus, or zoster sine herpete.

Immunocompromised patients may develop severe skin eruption with or without hemorrhage. Healing of the cutaneous lesions takes three times longer than in the general population. Patients develop high fever. Virus spreads to visceral organs causing hepatitis, pneumonitis, pancreatitis, small bowel obstruction and encephalitis. Bacterial superinfections including bacteremia can develop. Antiviral therapy significantly reduces morbidity and can reduce mortality.

Enzyme immunoassay (EIA), latex agglutination (LA), Indirect fluorescent antibody (IFA) and fluorescent antibody to membrane antigen assay (FAMA) are reliable in the immunocompetent patient but not necessarily in immunocompromised patients. The method of polymerase chain reaction (PCR) in the VZV diagnosis is highly sensitive, specific and it has become very important in the laboratory diagnostics of VZV.

#### **INTENDED USE**

Kit **VZV Real-TM** is a test for the qualitative detection of *Varicella Zoster virus* in the whole blood, liquor, swabs, urine, prostatic liquid and other biological materials.

#### PRINCIPLE OF ASSAY

Kit **VZV Real-TM** is based on two major processes: isolation of DNA from specimens and Real Time amplification. VZV DNA is extracted from the specimens, amplified using Real-Time amplification and detected fluorescent reporter dye probes specific for VZV DNA and Internal Control. Internal Control (IC) serves as an amplification control for each individually processed specimen and to identify possible reaction inhibition. IC is detected in a channel other than the VZV.

#### **MATERIALS PROVIDED**

#### Module No.1: Real Time PCR kit (V61-50FRT)

Part N° 2 – "VZV Real-TM": Real Time amplification

- PCR-mix-1-FRT, 0,6 ml;
- PCR-Buffer-FRT, 0,3 ml;
- TaqF Polymerase, 0,03 ml;
- Pos VZV C+, 0,1 ml;
- Negative Control C-, 2 x 0,5 ml;\*
- Internal Control IC, 0,6 ml;\*\*
- DNA-buffer, 0,5 ml;

Contains reagents for 55 tests.

# Module No.2: Complete Real Time PCR test with DNA purification kit (TV61-50FRT)

Part N° 1 – "DNA/RNA Prep": Sample preparation

- Lysis Sol, 15 ml;
- Prec Sol, 20 ml;
- Washing Sol 3, 25,0 ml;
- Washing Sol 4, 10,0 ml
- RE-buffer, 4 x 1,2 ml;

Contains reagents for 50 extractions

#### Part N° 2 – "VZV Real-TM": Real Time amplification

- PCR-mix-1-FRT, 0,6 ml;
- PCR-Buffer-FRT, 0,3 ml;
- TaqF Polymerase, 0,03 ml;
- Pos VZV C+, 0,1 ml;
- Negative Control C-, 2 x 0,5 ml;\*
- Internal Control IC, 0,6 ml;\*\*
- DNA-buffer, 0,5 ml;

Contains reagents for 55 tests.

\*must be used in the isolation procedure as Negative Control of Extraction.

\*\*add 10 µl of Internal Control during the DNA isolation directly to the sample/lysis mixture (see DNA/RNA Prep
|REF| K-2-9 protocol).

#### MATERIALS REQUIRED BUT NOT PROVIDED

#### **Zone 1: sample preparation:**

- DNA extraction kit (Module No. 1)
- Biological cabinet
- Desktop microcentrifuge for "eppendorf" type tubes
- Dry heat block
- Vortex mixer
- Pipettes
- Sterile pipette tips with filters
- 1,5 ml polypropylene sterile tubes
- Biohazard waste container
- · Refrigerator, freezer

# **Zone 2: Real Time amplification:**

- Real Time Thermal cycler
- Reaction tubes
- Workstation
- Pipettes (adjustable)
- Sterile pipette tips with filters
- Desktop centrifuge with rotor for 1,5/2,0 ml tubes
- Vortex mixer
- Freezer, refrigerator

#### STORAGE INSTRUCTIONS

**VZV Real-TM** must be stored at -20°C. DNA/RNA Prep must be stored at +2-8°C. **DNA/RNA Prep** must be stored at 2-8°C. The kits can be shipped at 2-8°C but should be stored at 2-8°C and -20°C immediately on receipt.

#### **STABILITY**

**VZV Real-TM** is stable up to the expiration date indicated on the kit label. The product will maintain performance through the control date printed on the label. Exposure to light, heat or humidity may affect the shelf life of some of the kit components and should be avoided. Repeated thawing and freezing of these reagents should be avoided, as this may reduce the sensitivity.

#### **QUALITY CONTROL**

In accordance with Sacace's ISO 13485-Certified Quality Management System, each lot is tested against predetermined specifications to ensure consistent product quality.

#### **WARNINGS AND PRECAUTIONS**



# In Vitro Diagnostic Medical Device

For In Vitro Diagnostic Use Only

The user should always pay attention to the following:

- Component Lysis Sol contains guanidine thiocyanate. Guanidine thiocyanate is harmful if inhaled, or comes into contact with skin or if swallowed. Contact with acid releases toxic gas. (Xn; R: 20/21/22-36/38; S: 16/36/37/39)\*.
- Component Prec Sol contains 2-propanol: flammable. Irritant. (R10-36-37-38-67, S7-16-24/25-26). In case of contact with eyes, rinse immediately with plenty of water and seek medical advice\*;
- Use sterile pipette tips with aerosol barriers and use new tip for every procedure.
- Store extracted positive material (samples, controls and amplicons) away from all other reagents and add it to the reaction mix in a separate area.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable gloves, laboratory coats and eye protection when handling specimens and reagents. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local authorities' regulations.
- Specimens should be considered potentially infectious and handled in a biological cabinet in accordance with appropriate biosafety practices.
- Clean and disinfect all sample or reagent spills using a disinfectant such as 0.5% sodium hypochlorite, or other suitable disinfectant.
- Avoid sample or reagent contact with the skin, eyes, and mucous membranes. If skin, eyes, or mucous membranes come into contact, rinse immediately with water and seek medical advice immediately.
- Material Safety Data Sheets (MSDS) are available on request.
- Use of this product should be limited to personnel trained in the techniques of DNA amplification.

• The laboratory process must be one-directional, it should begin in the Extraction Area and then move to the Amplification and Detection Areas. Do not return samples, equipment and reagents to the area in which the previous step was performed.



Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

#### PRODUCT USE LIMITATIONS

All reagents may exclusively be used in in vitro diagnostics. Use of this product should be limited to personnel trained in the techniques of DNA amplification (UNI EN ISO 18113-2:2012). Strict compliance with the user manual is required for optimal PCR results. Attention should be paid to expiration dates printed on the box and labels of all components. Do not use a kit after its expiration date.

#### SAMPLE COLLECTION, STORAGE AND TRANSPORT

**VZV Real-TM** can analyze DNA extracted from:

- whole blood collected in either ACD or EDTA tubes;
- plasma;
- *liquor* stored in "Eppendorf" tube;
- *tissue:* 1,0 gr homogenized with mechanical homogenizer or scalpel, glass sticks, teflon pestles and dissolved in 1,0 ml of saline water or PBS sterile. Vortex vigorously and incubate 30 min at room temperature. Transfer the supernatant into a new 1,5 ml tube;
- prostatic liquid stored in "Eppendorf" tube;
- seminal liquid: transfer about 30 μl of seminal liquid to a polypropylene tube (1,5 ml) and add 70 μl of sterile saline solution;
- swabs: insert the swab into the nuclease-free 1,5 ml tube and add 0,2 mL of Transport medium. Vigorously agitate swabs in medium for 15-20 sec.

Specimens can be stored at +2-8°C for no longer than 48 hours, or frozen at -20°C to -80°C. Transportation of clinical specimens must comply with country, federal, state and local regulations for the transport of etiologic agents.

<sup>\*</sup> Only for Module No.2

#### **DNA ISOLATION**

Any commercial RNA/DNA isolation kit, if IVD-CE validated for the specimen types indicated herein at the "SAMPLE COLLECTION, STORAGE AND TRANSPORT" paragraph, could be used.

Sacace Biotechnologies recommends to use the following kit:

- ⇒ DNA/RNA Prep (Sacace, REF K-2-9);
- ⇒ SaMag Viral Nucleic Acids Extraction kit (Sacace, REF SM003) for plasma, liquor;
- ⇒ SaMag STD DNA Extraction kit (Sacace, REF SM007) for seminal liquid, swabs.

Please carry out DNA extraction according to the manufacture's instruction. Add 10 µl of Internal Control during DNA isolation procedure directly to the sample/lysis mixture.

#### SPECIMEN AND REAGENT PREPARATION (reagents supplied with the module no.2)

- 1. Prepare required number of 1.5 ml disposable polypropylene micro centrifuge tubes including one tube for Negative Control of Extraction (**Negative Control**, **C-**).
- 2. Add to each tube 10 μl of IC (Internal Control) and 300 μl of Lysis Sol
- 3. Add **100 µl** of samples to the appropriate tubes using pipette tips with aerosol barriers.
- 4. Prepare Controls as follows:
  - o add 100 μl of Negative Control C- to the tube labeled Cneg
- 5. Vortex the tubes and incubate for 5 min at 65°C. Centrifuge for 7-10 sec.
- 6. Add **400 μl** of **Prec Sol** and mix by vortex. Centrifuge all tubes at 13,000 r/min for 5 min and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between the tubes.
- 7. Add **500 µl of Wash Sol 3** into each tube. Vortex vigorously to ensure pellet washing. Centrifuge all tubes at 13,000 r/min for 60 sec and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between the tubes.
- 8. Add 200 μl of Wash Sol 4 into each tube. Vortex vigorously to ensure pellet washing. Centrifuge all tubes at 13,000 r/min for 60 sec and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between the tubes.
- 9. Incubate all tubes with open caps at 65 °C for 5 min.
- 10. Resuspend the pellet in **50 μl of RE-buffer** (elution volume can be increased up to 90 μl). Incubate for 5 min at 65°C and vortex periodically.
- 11. Centrifuge the tubes at 13000g for 60 sec.

The supernatant contains RNA/DNA ready for amplification. If amplification is not performed the same day of extraction, the processed samples can be stored at 2-8°C for at maximum period of 5 days or frozen at - 20°/-80°C.

#### **PROTOCOL**:

- 1. Prepare required quantity of reaction tubes for samples (N) and controls (N+2).
- 2. Prepare in the new sterile tube for each sample 10\*(N+1) µI of PCR-mix-1-FRT, 5,0\*(N+1) of PCR-Buffer-FRT and 0,5\*(N+1) µI of TaqF DNA Polymerase. Vortex and centrifuge for 2-3 sec.
- 3. Add to each tube 15 µl of Reaction Mix and 10 µl of extracted DNA sample to appropriate tube. Mix by pipetting.
- 4. Prepare for each panel 2 controls:
  - add 10 µl of DNA-buffer to the tube labeled Amplification Negative Control;
  - add 10 μl of Positive VZV C+ to the tube labelled Amplification Positive Control;
- 5. Insert the tubes in the thermalcycler.

### **Amplification**

Create a temperature profile on your instrument as follows:

	Rotor-type instruments <sup>1</sup>			Plate- or modular type instruments <sup>2</sup>		
Step	Temperature °C	Time	Cycles	Temperature °C	Time	Cycles
Hold	95	15 min	1	95	15 min	1
Cycling 1	95	5 s		95	5 s	
	60	20 s	5	60	20 s	5
	72	15 s		72	15 s	
Cycling 2	95	5 s		95	5 s	
		20 s			30 s	
	60	fluorescent	40	60	fluorescent	40
		signal detection			signal detection	
	72	15 s		72	15 s	

Fluorescence is detected at the 2nd step of Cycling 2 stage (60 °C) in FAM/Green and JOE/Yellow/Hex/Cy3 fluorescence channels.

VZV is detected on the JOE(Yellow)/HEX/Cy3 channel, IC DNA on the FAM (Green) channel

<sup>&</sup>lt;sup>1</sup> For example Rotor-Gene<sup>™</sup> 3000/6000/Q (Corbett Research, Qiagen)
<sup>2</sup> For example, SaCycler-96<sup>™</sup> (Sacace), CFX96<sup>tm</sup>/iQ5<sup>™</sup> (BioRad); Mx3005P<sup>™</sup> (Agilent), ABI® 7300/7500/StepOne Real Time PCR (Applied Biosystems), SmartCycler® (Cepheid), LineGeneK® (Bioer)

# **INSTRUMENT SETTINGS**

# Rotor-type instruments (RotorGene 3000/6000, RotorGene Q)

Channel	Calibrate/Gain Optimisation	Threshold	More Settings/ Outlier Removal	Slope Correct
FAM/Green	from 5 Fl to 10 Fl	0.03	10 %	on
JOE/Yellow	from 4 FI to 8 FI	0.03	10 %	on

# Plate- or modular type instruments

The threshold line should cross only sigmoid curves of signal accumulation of positive samples and should not cross the baseline; otherwise, the threshold level should be raised. Set the threshold at a level where fluorescence curves are linear and do not cross curves of the negative samples.

#### **DATA ANALYSIS**

#### The fluorescent signal intensity is detected in two channels

- The signal from the VZV DNA amplification product is detected in the JOE(Yellow)/HEX/Cy3 channel:
- The signal from the Internal Control amplification product is detected in the FAM (Green) channel.

## Interpretation of results

The results are interpreted through the presence of crossing of fluorescence curve with the threshold line. To set threshold put the line at such level where curves of fluorescence are linear. Results are accepted as relevant if positive and negative controls of amplification and extraction are passed.

#### **Results for controls**

Control	Stage for control	Ct FAM (Green)	Ct JOE(Yellow)/HEX/Cy3	Interpretation
NCE	DNA isolation	POS	NEG	Valid result
NCA	Amplification	NEG	NEG	Valid result
C+ VZV	Amplification	NEG	POS	Valid result

- VZV DNA is detected in a sample if its Ct value is defined in the results grid in the JOE(Yellow)/HEX/Cy3 channel.
- VZV DNA is **not detected** in a sample if its Ct value is not defined in the results grid in the JOE(Yellow)/HEX/Cy3 channel (the fluorescence curve does not cross the threshold line) whereas the Ct value in the FAM (Green) channel is defined.
- The result of analysis is invalid if the Ct value is not defined in the results grid (the fluorescence curve does not cross the threshold line) in the FAM/Green channel. In this case, PCR should be repeated starting from the DNA extraction.

Boundary value of the cycle threshold, Ct

	Channel for	Ct boundary value		
Sample	fluorophore	Rotor-type instruments	Plate-type instruments	
C+	FAM/Green	30	30	
U <sup>+</sup>	JOE/Yellow/Hex/Cy3	30	30	
Clinical samples, C-	FAM/Green	30	30	

#### **QUALITY CONTROL PROCEDURE**

A defined quantity of Internal Control (IC) is introduced into each sample and control at the beginning of sample preparation procedure in order to control the extraction process of each individual sample and to identify possible reaction inhibition.

A negative control of extraction (NCE), negative amplification control (NCA), positive amplification control (C+) are required for every run to verify that the specimen preparation, the amplification and the detection steps are performed correctly.

If the controls are out of their expected range (see table Results for Controls), all of the specimens and controls from that run must be processed beginning from the sample preparation step.

#### **SPECIFICATIONS**

# **Analytical specificity**

Analytical specificity of the primers and probes was validated with 100 negative samples. They did not generate any signal with the specific for VZV primers and probes. The potential cross-reactivity of the kit **VZV Real-TM** was tested also against the group control listed below. The results of VZV were interpreted on the Joe channels while IC results (recombinant structure inserted in DNA of fago  $\lambda$ ) were interpreted on the Fam channel . The kit did not give any cross-reactivities with these pathogens. There were no nonspecific responses during examination of human DNA as well as DNA panel of the following microorganisms: *EBV*, *CMV*, *HSV 1 HSV 2*, *HHV6*, *Parvovirus B19*, *hRSV*, *Rubella*, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Streptococcus agalactiae* 

### Analytical sensitivity and reproducibility.

The analytical sensitivity of the **VZV Real-TM** kit was determined using the Standard DNA of the VZV. This Standard was serially diluted in the DNA-buffer.

The analytical sensitivity of the kit VZV Real-TM was not less than 500 copies/ml

#### **TROUBLESHOOTING**

- 1. Weak or no signal of the IC (FAM (Green) for the Negative Control of extraction.
  - The PCR was inhibited.
    - ⇒ Make sure that you use a recommended DNA extraction method and follow to the manufacturer's instructions.
    - ⇒ Re-centrifuge all the tubes before pipetting of the extracted DNA for 2 min at maximum speed (12000-16000 g) and take carefully supernatant. Don't disturb the pellet, sorbent inhibit reaction.
  - The reagents storage conditions didn't comply with the instructions.
    - ⇒ Check the storage conditions
  - The PCR conditions didn't comply with the instructions.
    - ⇒ Check the PCR conditions and select for the IC detection the fluorescence channel reported in the protocol.
  - The IC was not added to the sample during the pipetting of reagents.
    - ⇒ Make attention during the DNA extraction procedure.
- 2. Weak or no signal of the Positive Control.
  - The PCR conditions didn't comply with the instructions.
    - ⇒ Check the amplification protocol and select the fluorescence channel reported in the manual.
- 3. JOE(Yellow)/HEX/Cy3 signal with Negative Control of extraction.
  - Contamination during DNA extraction procedure. All samples results are invalid.
    - ⇒ Decontaminate all surfaces and instruments with sodium hypochlorite and ethanol.
    - ⇒ Use only filter tips during the extraction procedure. Change tips between tubes.
    - ⇒ Repeat the DNA extraction with the new set of reagents.
- 4. Any signal with Negative Control of PCR (DNA-buffer).
  - Contamination during PCR preparation procedure. All samples results are invalid.
    - ⇒ Decontaminate all surfaces and instruments with sodium hypochlorite and ethanol or special DNA decontamination reagents.
    - ⇒ Pipette the Positive control at last.
    - ⇒ Repeat the PCR preparation with the new set of reagents.

#### **KEY TO SYMBOLS USED**

REF	List Number		Caution!
LOT	Lot Number	$\sum_{i}$	Contains sufficient for <n> tests</n>
IVD	For <i>in Vitro</i> Diagnostic Use	VER	Version
	Store at	NCA	Negative Control of Amplification
	Manufacturer	NCE	Negative control of Extraction
i	Consult instructions for use	C+	Positive Control of Amplification
$\subseteq$	Expiration Date	IC	Internal Control



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Sacace™ VZV Real-TM

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